



## SM Agar

M763

### Intended Use

Recommended for cultivation and enumeration of microorganisms encountered in dairy industry.

### Composition\*\*

Ingredients	Gms / Litre
SM powder #	28.000
Tryptone	5.000
Yeast extract	2.500
Dextrose (Glucose)	1.000
Agar	15.000
Final pH ( at 25°C)	7.0±0.2

\*\*Formula adjusted, standardized to suit performance parameters

# Equivalent to Skim Milk powder

### Directions

Suspend 51.5 grams of in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C. Mix well and pour into sterile Petri plates.

### Principle And Interpretation

SM Agar is used for the demonstration of coagulation and proteolysis of casein (1). The medium is recommended by APHA (2) for cultivation and enumeration of microorganisms encountered in dairy industry (3). Addition of SM powder to any nutrient-rich medium creates favourable conditions for growth of organisms, which are encountered in milk. The number of bacteria isolated thus is more than the number of organisms isolated on a regular medium (4). Proteolytic bacteria hydrolyze casein to form soluble nitrogenous compounds indicated as clear zone surrounding the colonies. More clear zones are seen on milk agar if, the bacteria produce acid from fermentable carbohydrates in the medium.

Tryptone provides amino acids and other complex nitrogenous substances. Yeast extract supplies vitamin B complex. Addition of SM powder in the medium makes the conditions optimal for microorganisms encountered in milk. Glucose acts as the carbon source.

### Type of specimen

Dairy samples

### Specimen Collection and Handling

For Dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (3). After use, contaminated materials must be sterilized by autoclaving before discarding.

### Warning and Precautions:

In Vitro diagnostic Use only. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets

### Limitations:

1. Further biochemical identification is required for identification of species.
2. Some strains show less growth due to variable nutritional requirements,

## Quality Control

### Appearance

Cream to yellow homogeneous free flowing powder

### Gelling

Firm, comparable with 1.5% Agar gel

### Colour and Clarity of prepared medium

Off white coloured opaque gel forms in Petri plates

### Reaction

Reaction of 5.15% w/v aqueous solution at 25°C. pH : 7.0±0.2

### pH

6.80-7.20

### Cultural Response

M763: Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours.

Organism	Inoculum (CFU)	Growth	Recovery	Proteolytic activity
<i>Bacillus subtilis</i> ATCC 6633	50-100	good-luxuriant	≥70%	positive reaction, clear zone surrounding colonies
<i>Enterococcus faecalis</i> ATCC 29212	50-100	luxuriant	≥70%	negative reaction, no clear zone surrounding colonies
<i>Escherichia coli</i> ATCC 25922	50-100	good-luxuriant	≥70%	negative reaction, no clear zone surrounding colonies
<i>Proteus mirabilis</i> ATCC 25933	50-100	luxuriant	≥70%	positive reaction, clear zone surrounding colonies
<i>Pseudomonas aeruginosa</i> ATCC 27853	50-100	luxuriant	≥70%	positive reaction, clear zone surrounding colonies
<i>Serratia marcescens</i> ATCC 8100	50-100	luxuriant	≥70%	positive reaction, clear zone surrounding colonies

## Storage and Shelf Life

Store below 30°C in tightly closed container and the prepared medium at 2 - 8°C. Use before expiry date on the label.

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4. Terplan G. Rundfeldt, H. u. Zaadhof, K.J. Zur Eignung verschiedener Nährböden für die Bestimmung der Gesamtkeimzahl der Milch. - Arch. Lebensmittelhyg., 18; 9-11 (1967).
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3. Baird R.B., Eaton A.D., and Rice E.W., (Eds.), 2015, Standard Methods for the Examination of Water and Wastewater, 23rd ed., APHA, Washington, D.C.
4. Isenberg, H.D. Clinical Microbiology Procedures Handbook. 2<sup>nd</sup> Edition.
5. Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock, D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.
6. Lapage S., Shelton J. and Mitchell T., 1970, Methods in Microbiology', Norris J. and Ribbons D., (Eds.), Vol. 3A, Academic Press, London.
7. MacFaddin J. F., 2000, Biochemical Tests for Identification of Medical Bacteria, 3rd Ed., Lippincott, Williams and Wilkins, Baltimore.
8. Wehr H. M. and Frank J. H., 2004, Standard Methods for the Microbiological Examination of Dairy Products, 17th Ed., APHA Inc., Washington, D.C.

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