

## Moeller Decarboxylase HiVeg™ Broth Base / Broth MV393/MV687/MV688/MV689 with Lysine Hydrochloride / Broth with Ornithine Hydrochloride / Broth with Arginine Hydrochloride

Moeller Decarboxylase HiVeg Broth Base with the addition of appropriate L-amino acid, is used to differentiate bacteria on the basis of their ability to decarboxylate the specific amino acids.

### Composition \*\* :

	MV393	MV687	MV688	MV689
Ingredients	Grams/Litre	Grams/Litre	Grams/Litre	Grams/Litre
HiVeg peptone	5.00	5.00	5.00	5.00
HiVeg extract	5.00	5.00	5.00	5.00
Dextrose	0.50	0.50	0.50	0.50
Bromo cresol purple	0.01	0.01	0.01	0.01
Cresol red	0.005	0.005	0.005	0.005
Pyridoxal	0.005	0.005	0.005	0.005
L-Lysine hydrochloride	—	10.00	—	—
L-Ornithine hydrochloride	—	—	10.00	—
L-Arginine hydrochloride	—	—	—	10.00

Final pH (at 25°C) 6.0 ± 0.2

\*\* Formula adjusted, standardized to suit performance parameters.

### Directions :

Suspend 10.52 grams of MV393 and 20.52 grams of MV687/MV688/MV689 in 1000 ml distilled water. Add 10 gm. of L-Lysine, L-Arginine, L-Ornithine or other L-amino acids in MV393. When using DL-amino acids, use 2% concentration. Heat if necessary to dissolve the medium completely. When L-Ornithine is added, readjustment of the pH is required. Dispense in 5 ml amount in screw-capped tubes and sterilize by autoclaving at 15 lbs pressure (121°C) for 10 minutes. Suspend 20.52 grams of MV687 or MV688 or MV689 in 1000 ml distilled water and sterilize as mentioned above.

### Principle and Interpretation :

These media are used for differentiating gram-negative enteric bacilli on the basis of their ability to decarboxylate amino acids. The Decarboxylase Broth was introduced by Moeller for detecting the production of lysine and ornithine decarboxylase and arginine dihydrolase (1). Prior to Moeller's work, bacterial amino acid decarboxylases were studied by Gale (2) and Gale and Epps (3). These HiVeg media are prepared by replacing animal based peptones with vegetable peptones which are BSE/TSE risks free. Production of ornithine decarboxylase is a helpful criterion in differentiating *Klebsiella* and *Enterobacter* species. *Klebsiella* are non-motile and do not produce ornithine decarboxylase while *Enterobacter* are motile and produce ornithine decarboxylase except *Enterobacter agglomerans* (4).

These media contain HiVeg extract and HiVeg peptone which provide nitrogenous nutrients for the growth of bacteria. Dextrose is the fermentable carbohydrate and pyridoxal is the co-factor for the decarboxylase enzyme. Bromo cresol purple and cresol red are the pH indicators in

### Product Profile :

Vegetable based (Code MV)☉	Animal based (Code M)
MV393/MV687/MV688/MV689	M393/M687/M688/M689
HiVeg peptone	Peptic digest of animal tissue
HiVeg extract	Beef extract
<b>Recommended for</b>	: Differentiating bacteria on the basis of their ability to decarboxylate the amino acids.
<b>Reconstitution</b>	: (MV393) : 10.52 g/l
	: (MV687/MV688/MV689) : 20.52 g/l
<b>Quantity on preparation (500g)</b>	: (MV393) : 47.52 L
	: (100g) : (MV393) : 9.50 L
	: (100g) : (MV687/MV688/MV689) : 4.87 L
<b>pH (25°C)</b>	: 6.0 ± 0.2
<b>Supplement</b>	: (MV393) : Amino acids.
<b>Sterilization</b>	: 121°C / 10 minutes
<b>Storage</b>	: Dry Medium-Below 30°C, Prepared Medium 2 - 8°C.

the medium. When the medium is inoculated with the dextrose fermenting bacteria, the pH is lowered due to acid production which changes the colour of the indicator from purple to yellow. Acid produced stimulates decarboxylase enzyme. Decarboxylation of lysine yields cadaverine while putrescine is produced due to ornithine decarboxylation. Arginine is first hydrolyzed to ornithine which is then decarboxylated to form putrescine. Formation of these amines increases the pH of the medium, changing the colour of the indicator from yellow to purple. If the organisms do not produce the appropriate enzyme, the medium remains acidic, yellow in colour. Each isolate to be tested should also be inoculated into the basal medium tube lacking the amino acid.

Inoculated tubes must be protected from air with a layer of sterile mineral oil. Exposure to air may cause alkalization at the surface of the medium which makes the test invalid.

### Quality Control :

#### Appearance of powder

Greenish yellow coloured, homogeneous, free flowing powder.

#### Colour and Clarity

Purple coloured, clear solution without any precipitate.

#### Reaction

Reaction of 1.05% w/v of MV393 or 2.05% w/v of MV687/MV688 / MV689 aqueous solution is pH 6.0 ± 0.2 at 25°C.

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### Cultural Response

Cultural characteristics observed after inoculating tubes, overlaying with sterile mineral oil and incubating at 35 - 37°C for upto 4 days.

Organisms (ATCC)	Lysine	Arginine	Ornithine
<i>Citrobacter freundii</i> (8090)	—	±	±
<i>Enterobacter aerogenes</i> (13048)	+	—	+
<i>Escherichia coli</i> (25922)	±	±	±
<i>Klebsiella pneumoniae</i> (13883)	+	—	—
<i>Proteus vulgaris</i> (13315)	—	—	—
<i>Proteus mirabilis</i> (25933)	—	—	+
<i>Pseudomonas aeruginosa</i> (9027)	—	+	—
<i>Salmonella</i> serotype Paratyphi A	—	(+) or +	+
<i>Salmonella</i> serotype Typhi (6539)	+	(+) or —	—
<i>Shigella flexneri</i> (12022)	—	— or (+)	—
<i>Shigella sonnei</i> (25931)	—	±	+
<i>Shigella dysenteriae</i> (13313)	—	— or (+)	—
<i>Serratia marcescens</i> (8100)	+	—	+

Key : + = positive reaction, purple colour  
 — = negative reaction, yellow or no colour change  
 ± = variable  
 (+) = delayed positive reaction



**MV689 Moeller Decarboxylase HiVeg Broth with Arginine Hydrochloride**

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|----------------------------------|-------------------------------------|
| 1. Control                       | 6. <i>Proteus vulgaris</i>          |
| 2. <i>Citrobacter freundii</i>   | 7. <i>Pseudomonas aeruginosa</i>    |
| 3. <i>Enterobacter aerogenes</i> | 8. <i>Salmonella</i> serotype Typhi |
| 4. <i>Escherichia coli</i>       | 9. <i>Shigella flexneri</i>         |
| 5. <i>Klebsiella pneumoniae</i>  |                                     |

### References :

1. Moeller V., 1955, Acta Pathol. Microbiol. Scand. 36:158.
2. Gale G. F., 1940, Biochem. J., 34:392.
3. Gale and Epps, 1943, Nature, 152:327.
4. MacFaddin J., 2000, Biochemical Tests for Identification of Medical Bacteria, 3<sup>rd</sup> ed., Williams and Wilkins, Baltimore.