

RS HiVeg™ Medium Base**MV576**

RS (Rimler-Shotts) HiVeg Medium is used for selective isolation, cultivation and presumptive identification of *Aeromonas hydrophila*.

Composition ** :

Ingredients	Grams/Litre
Yeast extract	3.0
Maltose	3.5
L-Cysteine hydrochloride	0.3
L-Lysine hydrochloride	5.0
L-Ornithine hydrochloride	6.5
Sodium thiosulphate	6.8
Ferric ammonium citrate	0.8
Synthetic detergent No. III	1.0
Sodium chloride	5.0
Bromo thymol blue	0.03
Agar	13.5

Final pH (at 25°C) 7.0 ± 0.2

** Formula adjusted, standardized to suit performance parameters.

Directions :

Suspend 45.43 grams in 990 ml distilled water. Heat to boiling to dissolve the medium completely. DO NOT AUTOCLAVE. Cool to 45°- 50°C and aseptically add Novobiocin Supplement (FD096). Mix well before pouring into sterile petriplates.

Principle and Interpretation :

This medium is prepared by using synthetic detergent No. III instead of sodium deoxycholate which makes the medium free of BSE/TSE risks. This medium is the modification of RS Medium developed by Rimler and Shotts for rapid isolation and identification of *Aeromonas hydrophila*(1). RS medium shows differentiation characteristics depending upon the biochemical reactions. The organisms that ferment maltose are seen as yellow colonies. The organisms that ferment maltose and are H₂S producers give yellow colour colonies with a black center. The third colonial type give greenish - yellow to green colonies which indicate decarboxylation of lysine or ornithine or both. The colonies which are greenish - yellow to green with black center indicate the decarboxylation of both amino acids and H₂S production.

The balance of ingredients provide a nutrient base and chemophysical stability for the medium. Hydrogen sulfide production depends upon the utilization of sodium thiosulphate or L- cysteine hydrochloride, or both, with ferric ammonium citrate being utilized to help visualize. Synthetic detergent No. III and Novobiocin helps in inhibiting the gram positive organisms and *Vibrio* species. Maltose fermentation is indicated by bromo thymol blue. Incubation should be done at 37°C and the results should be seen after 20 hours but not after 24 hours, as plates observed after 26 hours demonstrate a slow reversion of the fermentation reaction toward a basic reaction.

Product Profile :

Vegetable based (Code MV)©	Animal based (Code M)
MV576 Synthetic detergent No. III	M576 Sodium deoxycholate
Recommended for	: Selective isolation, cultivation and presumptive identification of <i>Aeromonas hydrophila</i> .
Reconstitution	: 45.43 g/l
Quantity on preparation (500g)	: 11.00 L
pH (25°C)	: 7.0 ± 0.2
Supplement	: Novobiocin Supplement (FD096)
Sterilization	: Boiling. (DO NOT AUTOCLAVE)
Storage : Dry Medium - Below 30°C, Prepared Medium 2 - 8°C.	

Quality Control :**Appearance of powder**

Light green coloured, homogeneous, free flowing powder.

Gelling

Firm, comparable with 1.35% Agar gel.

Colour and Clarity

Dark green coloured, clear gel forms in petri plates.

Reaction

Reaction of 4.54% w/v aqueous solution is pH 7.0 ± 0.2 at 25°C

Cultural Response

Cultural characteristics observed after an incubation at 35-37°C for 24 hours on addition of Novobiocin supplement (FD096).

Organisms (ATCC)	Inoculum (CFU)	Recovery	Maltose fermentation	Lysine/Ornithine*	H ₂ S
<i>Aeromonas hydrophila</i> (7966)	10 ² -10 ³	>50%	+	-	-
<i>Citrobacter freundii</i> (8090)	10 ² -10 ³	>50%	-	V	+
<i>Escherichia coli</i> (25922)	10 ² -10 ³	>50%	-	V	-
<i>Proteus vulgaris</i> (13315)	10 ² -10 ³	>50%	+	-	+
<i>Salmonella</i> serotype Typhi (6539)	10 ² -10 ³	>50%	+	-	-

Key : Maltose fermentation : + = yellow coloured colonies
 Lysine and/or Ornithine : + = shades of greenish yellow to yellow coloured colonies
 *decarboxylation
 Hydrogen Sulphide (H₂S) production : + = black centered colonies
 - = Negative reaction
 V = Variable

References :

1. Shotts E. B. Jr. and Rimler R., 1973, Appl. Microbiol. , 26(4):550