

## Moxifloxacin MO (Part A: 240 - 0.01 µg & Part B: 32 - 0.005 µg)

MD033

Moxifloxacin HiComb™ MIC Strip is a rapid and reliable method for determining the antimicrobial susceptibility of different microorganisms against Moxifloxacin. This system provides a set of 16 different concentrations in gradient that can be easily used to deduce a functionally accurate the Minimum Inhibitory Concentration (MIC) in microgram levels.

### Composition

#### Part Levels in µg of Moxifloxacin

Part A: 240, 120, 60, 30, 10, 5, 0.1, 0.01

Part B: 32, 16, 8, 4, 0.1, 0.06, 0.03, 0.005

### Susceptibility Test Procedure:

1. Prepare plates with Mueller Hinton Agar (M173/M1084) for rapidly growing aerobic organisms as per Bauer-Kirby Method. For fastidious organisms such as *Streptococcus*, the Medium is supplemented with 5% sterile, defibrinated blood. For *Haemophilus* spp, Haemophilus Test Agar Base (M1259) with added supplement (FD117) and for *N. gonorrhoeae*, GC Agar Base (M434) with 1% defined growth supplement (FD025) are recommended respectively. The medium in the plates should be sterile and should have a depth of about 4 mm.
2. Inoculate 4-5 similar colonies with a wire, needle or loop to 5 ml Tryptone Soya Broth (M011) and incubate at 35-37°C for 2-8 hours until light to moderate turbidity develops. Compare the inoculum turbidity with that of standard 0.5 McFarland (prepared by mixing 0.5 ml of 1.175% barium chloride and 99.5 ml of 0.36N sulfuric acid). Dilute the inoculum or incubate further as necessary to attain comparative turbidity. Alternatively, the inoculum can be standardized by other appropriate optical method (0.08 - 0.13 OD turbid suspension at 625 nm)
3. Dip a sterile non-toxic cotton swab on a wooden applicator into the standardized inoculum and rotate the soaked swab firmly against the upper inside wall of the tube to express excess fluid. Streak the entire agar surface of the plate with the swab three times, turning the plate at 60° angle between each streaking. Allow the inoculum to dry for 5 - 15 minutes with lid in place.
4. Apply the strips using aseptic technique as follows
  - i. Pick up one strip by its handle with a sterile forceps
  - ii. Place the strip with its higher concentration facing the edge of plate and the markings on strip facing up wards.
  - iii. Press gently on the handle of the strip and assure that all the discs are in full contact with the medium.
  - iv. Place other strip on the opposite side of plate with higher concentration towards the edge of plate and lower concentration towards the centre.
  - v. Close the plate and invert to check whether all the discs are in full contact with the medium [6 Strips (3 of A and 3 of B) can be placed on a 200 mm plate with the markings facing upward]
5. Incubate immediately at 35 ± 2°C and examine after 16-18 hours or longer, if necessary. For fastidious organisms incubate at appropriate temperature and time.
6. The zone of inhibition will be in the form of an ellipse. MIC value would be the value at which the zone convenes the comb-like projections of the strips and not at the handle. If there is no zone of inhibition observed, report the MIC as greater than the highest concentration on the strip. If the zone of inhibition is below the lowest concentration then report the MIC as less than the lowest concentration.

### Principle:

Antimicrobial susceptibility testing (AST) of bacterial and fungal isolates is a common and important technique in most clinical laboratories. The results of these tests are used for selection of the most appropriate antimicrobial agent(s) for treatment against the infectious organisms. Till the 1950s, laboratories were lacking in the methodologies and equipments for the accurate determination of in vitro responses of organisms to antimicrobial agents. Bauer et al (1) began the development of standardized methods for antimicrobial susceptibility testing, using disc diffusion system. However the susceptibility results may not always correlate with the patient's response to therapy. The response of an infected patient to antimicrobial agent(s) is a complex interrelationship of host responses, drug dynamics and microbial activity. Antimicrobial susceptibility tests are either quantitative or

qualitative. Minimum Inhibitory Concentration (MIC) determination is a Quantitative method. Determining MIC using a conventional broth dilution method is a tedious and time consuming process.

Hence Gradient diffusion susceptibility testing method is accepted widely and is in routine use for a long time. HiComb is also based on innovative gradient-based technique. This system is developed using dry chemistry technology and consists of two comb shaped strips made of an inert material with 8 extensions that carry the discs of 4 mm, resembling the 'tooth' of a comb. Towards the stem of the strip, MIC reading scale in µg/ml is given along with the HiMedia code. A defined concentration of antibiotic is loaded on each of the disc so as to form a gradient when placed on agar plate. HiComb based on the principle of dilution and diffusion, consists of a gradient that cover a continuous range across 16 two-fold dilutions as per the conventional MIC method. When applied to the agar surface, the antibiotic instantaneously diffuses into the surrounding medium in high to low amounts from one end of the strip to the other. The gradient remains stable after diffusion, and the zone of inhibition created takes the form of ellipse. The MIC value would be the value at which the zone convenes the comb-like projections of the strips and not at the handle. The MIC value obtained from this system can be compared to the standardized CLSI procedure (2).

HiComb is advantageous for routine use as it is simple to set up and perform, and is rapid than any agar or broth dilution method. The test has special advantage to study the resistance surveillance. The wide concentration gradients of these tests cover the MIC ranges of susceptibility of a wide variety of pathogens and allow both low level and high-level resistance detection.

### Interpretation:

Use following interpretive criteria for susceptibility categorization.

Antimicrobial agent	Interpretative criteria for	Sensitive ≤	Intermediate	Resistant ≥
Moxifloxacin	<i>Staphylococcus</i>	0.5	1	2
	<i>S.pneumoniae</i>	1	2	4
	<i>Haemophilus</i>	1	-	-

### Quality Control:

**Appearance:** Comb like inert strips having 16 different concentrations of Moxifloxacin (8 on each strip) impregnated on the circular filter paper discs having diameter of 4mm with the concentrations printed on the strips.

**Cultural response:** Zone of inhibition in form of ellipse is observed on Mueller Hinton Agar (M173) after incubation at 35-37°C for 18-24 hours for standard cultures.

Organisms (ATCC)	Std. MIC range (µg)
<i>S.aureus</i> (29213)	0.015-0.12
<i>E.faecalis</i> (29212)	0.06-0.5
<i>E. coli</i> (25922)	0.008-0.06
<i>P.aeruginosa</i> (27853)	1-8

### Storage and Shelf-life:

Strips should always be stored at -20°C to +8°C under dry conditions, along with the dessicator provided in individual pack. Use before expiry date on the label.

**References:**

1. Bauer, Kirby, Sherris and Turck, 1966, Am. J. Clin. Path., 45: 493
2. Performance standards of Antimicrobial Disc Susceptibility Tests, CLSI Vol. 32 No.3, Jan 2012.

**Note:**

1. This strip is INTENDED ONLY FOR AGAR DIFFUSION METHOD AND NOT FOR BROTH DILUTION METHOD.
2. HiComb should be used for *in vitro* diagnostic purpose only and aseptic procedures and precautions should be used when handling bacterial samples throughout the testing procedure.
3. Before using HiComb strips, ensure that the strip is at Room temperature or 25°C.
4. Pick the HiComb strip by its handle.
5. If accidentally dropped on to a dry surface, pick the strip up and use it.
6. If dropped in, an incorrect position, leave the strip there and use the other region of the plate.
7. When applying strips be steady. Do not move the strip once in contact with the agar surface, since the antibiotic instantaneously diffuse on contact with agar.
8. For fastidious organisms or very susceptible species, use only 4 to 5 strips on a 150 mm plate and one on a 90 mm plate.

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