

**MBRE014**

**Sph I**

**Components**

Reagents provided	MBRE014			
	250 Units	500 Units	1000 Units	5000 Units
Sph I	50 µl	100 µl	200 µl	1 ml
10X HiBuffer DB	1.25 ml	2.5 ml	5 ml	25 ml
Diluent E Buffer	625 µl	1.25 ml	2.5 ml	12.5 ml

**NOTE: BSA included in all Reaction Buffer**

**Source:** A *E. coli* strain that carries the Sph I gene from *Streptomyces phaeochromogenes*

**Recognition Sequence:**



**Concentration:** 5 U/µl

**Unit Definition:**

1 u is defined as the amount of enzyme that is required to digest 1µg of DNA in 1 hour at 37°C in 50 µl of assay buffer.

Enzyme	Optimum reaction temperature (°C)	Thermal Inactivation (°C)	% activity of Buffers				
			H1	H2	H3	H4	H5
Sph I	37	65	75	75	50	75	75

**Reaction Buffer:**

**10X HiBuffer DB:**

25mM Tris –acetate (pH 7.6 at 30°C), 10 mM Mg-acetate, 100 mM K-acetate, 7mM 2-mercaptoethanol and 50 µg /ml BSA.

**NOTE:** Incubate at 37°C

**Storage Buffer:**

10mM Tris-HCl (pH 7.5), 300mM NaCl, 0.1mM EDTA, 7mM 2-mercaptoethanol, 200µg/ml BSA and 50% glycerol. Store at –20°C .

**NOTE: 10X HiBuffer DB is provided for double digestion.**

**Quality Control Assays:**

**Ligation / Recutting Assay:**

After 10-fold over digestion with Sph I, more than 90% of the DNA fragments can be ligated and recut.

**Over digestion Assay:**

An unaltered banding pattern was observed after 1µg of DNA was digested with 20U of Sph I for 16 hours at 37°C.

**Example of Digestion conditions:**

- Enzyme concentration : 1 Unit
- Lambda 0.3 µg/µl : 3.33 µl (1 µg DNA)
- 10X HiBuffer DB : 5 µl
- Nuclease free water : upto 50 µl

**Note:**

- Total reaction volume is dependent on the experiment
- The amount of enzyme to be used is dependent on the DNA template
- For plasmid DNA, 5-10X more enzyme is required
- High enzyme concentration may result in **Star activity**

**Storage conditions:** Sph I should be stored at -20°C.