

## RPMI 1640

With L- Glutamine and 25mM HEPES buffer  
Without Sodium bicarbonate

**Product Code: AT060**

### Product Description :

Roswell Park Memorial Institute (RPMI) media are a series of media developed by Moore et al for the culture of human normal and neoplastic cells in vitro. RPMI-1640 is the most commonly used medium in the series. A modification of McCoy's 5a medium, the medium was specifically designed to support the growth of human lymphoblastoid cells in suspension culture. Presently the medium is extensively used for a wide range of anchorage dependant cell lines. The medium needs to be supplemented with 5-20% fetal bovine serum. The medium is also known to support growth of cells in the absence of serum.

AT060 is modified RPMI-1640 medium supplemented with 25mM HEPES buffer and L-glutamine. HEPES, a zwitterionic buffer having a pKa of 7.3 at 37°C prevents the initial rise in pH that tends to occur at the initiation of a culture and increases the buffering capacity of the medium. Users are advised to review the literature for recommendations regarding medium supplementation and physiological growth requirements specific for different cell lines.

### Composition :

| Ingredients                        | mg/L     |
|------------------------------------|----------|
| <b>INORGANIC SALTS</b>             |          |
| Calcium nitrate tetrahydrate       | 100.000  |
| Magnesium sulphate anhydrous       | 48.840   |
| Potassium chloride                 | 400.000  |
| Sodium chloride                    | 6000.000 |
| Sodium phosphate dibasic anhydrous | 800.000  |
| <b>AMINO ACIDS</b>                 |          |
| Glycine                            | 10.000   |
| L-Arginine hydrochloride           | 241.000  |
| L-Asparagine                       | 50.000   |
| L-Aspartic acid                    | 20.000   |
| L-Cystine dihydrochloride          | 65.200   |
| L-Glutamic acid                    | 20.000   |
| L-Glutamine                        | 300.000  |
| L-Histidine hydrochloride          | 20.960   |

|                          |        |
|--------------------------|--------|
| L-Hydroxyproline         | 20.000 |
| L-Isoleucine             | 50.000 |
| L-Leucine                | 50.000 |
| L-Lysine hydrochloride   | 40.000 |
| L-Methionine             | 15.000 |
| L-Phenylalanine          | 15.000 |
| L-Proline                | 20.000 |
| L-Serine                 | 30.000 |
| L-Threonine (Allo free)  | 20.000 |
| L-Tryptophan             | 5.000  |
| L-Tyrosine disodium salt | 28.830 |
| L-Valine                 | 20.000 |

#### VITAMINS

|                             |        |
|-----------------------------|--------|
| Choline chloride            | 3.000  |
| D-Biotin                    | 0.200  |
| D-Ca-Pantothenate           | 0.250  |
| Folic acid                  | 1.000  |
| Niacinamide                 | 1.000  |
| Pyridoxine hydrochloride    | 1.000  |
| Riboflavin                  | 0.200  |
| Thiamine hydrochloride      | 1.000  |
| Vitamin B12                 | 0.005  |
| i-Inositol                  | 35.000 |
| p-Amino benzoic acid (PABA) | 1.000  |

#### OTHERS

|                        |          |
|------------------------|----------|
| D-Glucose              | 2000.000 |
| Glutathione reduced    | 1.000    |
| HEPES Buffer           | 5958.000 |
| Phenol red sodium salt | 5.300    |

### Directions :

1. Suspend 16.4gms in 900ml tissue culture grade water with constant, gentle stirring until the powder is completely dissolved. Do not heat the water.
2. Add 2.0gms of Sodium bicarbonate powder (TC230) or 26.67ml of 7.5% Sodium bicarbonate solution (TCL013) for 1 litre of medium and stir until dissolved.
3. Adjust the pH to 0.2-0.3 pH units below the desired pH using 1N HCl or 1N NaOH since the pH tends to rise during filtration.

4. Make up the final volume to 1000ml with tissue culture grade water.
5. Sterilize the medium immediately by filtering through a sterile membrane filter with a porosity of 0.22 micron or less, using positive pressure rather than vacuum to minimize the loss of carbon dioxide.
6. Aseptically add sterile supplements as required and dispense the desired amount of sterile medium into sterile containers.
7. Store liquid medium at 2-8°C and in dark till use.

### **Material required but not provided :**

Tissue culture grade water (TCL010)  
Sodium bicarbonate (TC230)  
Sodium bicarbonate solution, 7.5% (TCL013)  
1N Hydrochloric acid (TCL003)  
1N Sodium hydroxide (TCL002)  
Foetal bovine serum (RM1112/RM10432)

### **Quality Control:**

#### **Appearance**

Off-white to Creamish white, homogenous powder.

#### **Solubility**

Clear solution at 16.4gms/L.

#### **pH without Sodium Bicarbonate**

6.40 -7.00

#### **pH with Sodium Bicarbonate**

6.70 -7.30

#### **Osmolality without Sodium Bicarbonate**

280.00 -320.00

#### **Osmolality with Sodium Bicarbonate**

320.00 -360.00

#### **Cultural Response**

The growth promotion capacity of the medium is assessed qualitatively by analyzing the cells for the morphology and quantitatively by estimating the cell counts and comparing it with a control medium through minimum three subcultures.

#### **Endotoxin Content**

NMT 5EU/ml

### **Storage and Shelf Life:**

1. All the powdered media and prepared liquid culture media should be stored at 2-8°C. Use before the expiry date. In spite

of above recommended storage condition, certain powdered medium may show some signs of deterioration /degradation in certain instances. This can be indicated by change in colour, change in appearance and presence of particulate matter and haziness after dissolution.

2. Preparation of concentrated medium is not recommended since free base amino acids and salt complexes having low solubility may precipitate in concentrated medium.

3. pH and sodium bicarbonate concentration of the prepared medium are critical factors affecting cell growth. This is also influenced by amount of medium and volume of culture vessel used (surface to volume ratio). For example, in large bottles, such as Roux bottles pH tends to rise perceptibly as significant volume of carbon dioxide is released. Therefore, optimal conditions of pH, sodium bicarbonate concentration, surface to volume ratio must be determined for each cell type. We recommend stringent monitoring of pH. If needed, pH can be adjusted by using sterile 1N HCl or 1N NaOH or by bubbling in carbon dioxide.

4. If required, supplements can be added to the medium prior to or after filter sterilization observing sterility precautions. Shelf life of the medium will depend on the nature of supplement added to the medium.

Revision : 1 / 2011



#### **Disclaimer :**

User must ensure suitability of the product(s) in their application prior to use. Products conform solely to the information contained in this and other related HiMedia™ publications. The information contained in this publication is based on our research and development work and is to the best of our knowledge true and accurate. HiMedia™ Laboratories Pvt Ltd reserves the right to make changes to specifications and information related to the products at any time. Products are not intended for human or animal diagnostic or therapeutic use but for laboratory, research or further manufacturing use only, unless otherwise specified. Statements contained herein should not be considered as a warranty of any kind, expressed or implied, and no liability is accepted for infringement of any patents.