Soyabean Casein Digest Agar Plate w/ Lecithin and Polysorbate 80 MP449G
(Tryptone Soya Agar Plate w/ Lecithin & Polysorbate 80) (γ - irradiated)

**Intended use**
Recommended for determining efficiency of sanitization of containers, equipment, surfaces, water miscible cosmetics etc.

**Composition**

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Gms / Litre</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tryptone</td>
<td>15.000</td>
</tr>
<tr>
<td>Soya Peptone</td>
<td>5.000</td>
</tr>
<tr>
<td>Sodium chloride</td>
<td>5.000</td>
</tr>
<tr>
<td>Lecithin</td>
<td>0.700</td>
</tr>
<tr>
<td>Polysorbate 80 (Tween 80)</td>
<td>5.000</td>
</tr>
<tr>
<td>Agar</td>
<td>15.000</td>
</tr>
<tr>
<td>Final pH ( at 25°C)</td>
<td>7.3±0.2</td>
</tr>
</tbody>
</table>

**Directions**
Either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically on the plate.

**Principle And Interpretation**
Tryptone Soya Agar with Lecithin and Polysorbate 80 is used in RODAC (Replicate Organism Detection and Counting) plates for the detection and enumeration of microorganisms present on surfaces of sanitary importances.
Tryptone and Soya peptone provide nitrogenous compounds and other nutrients essential for microbial replication. Lecithin and polysorbate 80 (Tween 80) are neutralizers reported to inactivate residual disinfectants from where the sample is collected. Lecithin neutralizes quaternary ammonium compounds and polysorbate 80 neutralizes phenolic disinfectants, hexachlorophene, formalin and with lecithin ethanol.
Collection of samples from areas before and after the treatment with disinfectant evaluates cleaning procedures in environmental sanitation. The presence and number of microorganisms is determined by the appearance of colonies on the agar surface. After counting the colonies, carry out biochemical testing for identification.

**Type of specimen**
Swabs of containers, Equipment surfaces, Water miscible cosmetics etc.

**Specimen Collection and Handling**
For swabs of containers, equipment surfaces, water miscible cosmetics samples follow appropriate techniques for handling specimens as per established guidelines.
After use, contaminated materials must be sterilized by autoclaving before discarding.

**Warning and Precautions**
Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling specimens. Safety guidelines may be referred in individual safety data sheets.

**Limitations**
1. Individual strain of a microorganism may have unique growth requirements with respect to nutrients and physical conditions. Based on which the growth pattern of each varies on a medium and some even may display significant delay in development.
3) Each lot of the medium has been tested for the organisms specified on the COA. It is recommended to users to validate the medium for any specific microorganism other than mentioned in the COA based on the user’s unique requirement.

4) It is recommended to store the plates ta 24-30°C to avoid minimum condensation.

**Performance and Evaluation**
Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

**Quality Control**
Appearance
Sterile Soybean Casein Digest Agar Plate w/ Lecithin and Polysorbate 80 (γ - irradiated) in 90 mm disposable plates.

**Colour of medium**
Amber coloured medium

**Quantity of medium**
30 ml of medium in 90 mm disposable plates.

**Reaction**
7.10-7.50

**Sterility Test**
Passes release criteria

**Dose of irradiation (Kgy)**
13.00-20.00

**Cultural Response**
Growth Promotion was observed after an incubation at 30-35°C for 18-24 hours for bacteria and for fungus <=5 days.

**Recovery rate**
Recovery rate is considered 100% for bacterial growth on Blood Agar and fungal growth on Sabouraud Dextrose Agar.

**Growth promoting properties**
Growth of microorganism comparable to that previously obtained with previously tested and approved lot of medium occurs at the specified temperature for not more than the shortest period of time specified inoculating <=100 cfu (at 30-35°C for 18 hours).

<table>
<thead>
<tr>
<th>Organism</th>
<th>Inoculum (CFU)</th>
<th>Growth</th>
<th>Observed Lot value (CFU)</th>
<th>Recovery</th>
<th>Incubation temperature</th>
<th>Incubation period</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bacillus subtilis subsp. spizizenii ATCC 6633 (00003*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Staphylococcus aureus subsp. aureus ATCC 25923 (00034*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Staphylococcus aureus subsp. aureus ATCC 6538 (00032*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Escherichia coli ATCC 25922 (00013*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Escherichia coli ATCC 8739 (00012*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Escherichia coli NCTC 9002</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa ATCC 27853 (00025*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Pseudomonas aeruginosa ATCC 9027 (00026*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Salmonella Abony NCTC 6017</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Micrococcus luteus ATCC 9341</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Streptococcus pneumoniae ATCC 6305</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
<tr>
<td>Salmonella Typhimurium ATCC 14028 (00031*)</td>
<td>50-100</td>
<td>luxuriant</td>
<td>35-100</td>
<td>&gt;=70 %</td>
<td>30-35 °C</td>
<td>18-24 hrs</td>
</tr>
</tbody>
</table>

Please refer disclaimer Overleaf.
**Technical Data**

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**Storage and Shelf Life**

On receipt store between 20-30°C Use before expiry date on the label.

Product performance is best if used within stated expiry period.

**Disposal**

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with sample must be decontaminated and disposed of in accordance with current laboratory techniques (4,5).

**Reference**

2. Favero (Chairm), 1967, Biological Contamination Control Committee, a state of the art report., Am. Assoc. for contamination control.

**Revision:** 02 / 2020

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