**MBPCR025  Listeria monocytogenes Detection Kit (Real-Time)**

**Description:**

*Listeria monocytogenes* is a Gram-positive intracellular foodborne pathogen, most commonly isolated from raw meat, unpasteurized milk and dairy products, vegetables and seafood. This pathogen is known to cause Listeriosis, a type of food poisoning. In infected individuals, listeriosis is characterized by meningitis, encephalitis and septicemia. Less commonly, these infections will result in cutaneous lesions and flu-like symptoms. The mortality rate for those contracting listeriosis is approximately 20%.

Due to its ability to survive high and low temperatures as well as low pH, it can resist various food processing technologies and grow even at storage temperatures.

A number of molecular and cellular determinants of virulence have been identified for this intracellular pathogen and there is evidence of polymorphism among different strains of *L. monocytogenes*. Therefore, all *L. monocytogenes* strains are considered to be pathogenic.

Specific and faster methods for detection of foodborne pathogens, such as Real–time PCR, are the need of an hour. These techniques help to detect targeted pathogens quickly; this early and precise detection helps to take further actions.

**NOTE:** The *Listeria monocytogenes* Detection Kit (Real-Time) is for *in vitro* use only.

**Principle**

*Listeria monocytogenes* has several important virulence markers. One of the most important markers is **p60** which is encoded by *invasive associated protein* *(iap) gene which plays a vital role in intestinal invasion. This gene is responsible for identification of *Listeria monocytogenes* giving an amplification of 131 bp product.*

Real-time polymerase chain reaction, also called quantitative Polymerase Chain Reaction (qPCR) or kinetic Polymerase Chain Reaction, is a laboratory technique based on the principle of Polymerase Chain Reaction. This technique is used to amplify and simultaneously quantitate a targeted DNA sequence. Real-time PCR systems based on SYBr Green assays have increasingly been used for accurate, reliable detection and quantitation of various food-borne pathogens. HiMedia's *Listeria monocytogenes Detection kit (Real-time), is one such SYBr green based qPCR technique which allows amplification of iap gene.*
A. Diagrammatic representation of preferential binding of SYBr Green Dye to specific DNA fragments in real-time PCR.

Features:
- Fast and simple
- Sensitive and specific results
- Guaranteed reproducible results
- Rapid detection of all relevant clinical pathogens

Kit Contents:
The provided PCR kit contains:

<table>
<thead>
<tr>
<th>Components</th>
<th>Reagents provided for 10R (reactions)*</th>
<th>Reagents provided for 25R (reactions)*</th>
<th>Reagents provided for 50R (reactions)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hi-SYBr master mix (2X master mix containing SYBr Green, Assay buffer, Taq Polymerase, MgCl₂, dNTPs) (MBT074)</td>
<td>150 µl</td>
<td>400 µl</td>
<td>700 µl</td>
</tr>
<tr>
<td>Primer Mix</td>
<td>25 µl</td>
<td>60 µl</td>
<td>120 µl</td>
</tr>
<tr>
<td>Nuclease free water (ML065)</td>
<td>1 ml</td>
<td>2 ml</td>
<td>4 ml</td>
</tr>
</tbody>
</table>

* For a 20µl PCR reaction

General Preparation Instructions:
- Before use all PCR components should be completely thawed on ice (4°C).
- Perform the amplification reactions in a clean area, preferably in a biosafety cabinet.
Use of aerosol barrier pipette tips is recommended to reduce contamination risks from extraneous DNA templates.

Extract and store positive control sample (if used) separately from all other reagents to avoid contamination and add it to the reaction mix in a separate area.

**Sampling and Handling:**

**Sample Preparation:**

Various food, clinical and environmental samples and cultured bacteria are routinely examined.

For extraction and purification of high yield and pure bacterial DNA, perform the nucleic acid purification using HiMedia’s HiPurA™ Bacterial Genomic DNA Purification Kit (MB505) as instructed in the protocol.

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**Flow Chart for setting up PCR Reaction**

Add 10µl Hi-SYBr master mix (MBT074) in a PCR tube

↓

Add 2 µl the Primer mix (Final concentration 10 pmoles provided)

↓

Add 1-2 µl template DNA (upto 50 ng of extracted DNA)

↓

Add nuclease free water (ML065) to make the final volume to 20 µl

↓

Centrifuge the tube briefly at 6000 rpm for about 10 seconds.

↓

Place the tubes in real-time PCR machine and set the recommended PCR program (mentioned below)

↓

Interpret the data from the amplification plot (observe the Ct values)

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**Recommended PCR program:**

1. Initial denaturation : 95°C for 5 minutes

2. Cycling Parameters (No. of cycles: 30)
   - Denaturation : 95°C for 45 seconds
   - Annealing : 60°C for 45 seconds
   - Extension : 72°C for 45 seconds
3. Final Extension : 72°C for 8 minutes.

**Amplification Data:**

![Amplification graph](image)

<table>
<thead>
<tr>
<th>Sr. No.</th>
<th>Sample</th>
<th>C&lt;sub&gt;t&lt;/sub&gt; value</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Negative control</td>
<td>N/A</td>
</tr>
<tr>
<td>2</td>
<td>1 µl of template DNA (amplicon of <em>L. monocytogenes</em>)</td>
<td>13.36</td>
</tr>
<tr>
<td>3</td>
<td>1 µl of template (amplicon of <em>L. monocytogenes</em>)</td>
<td>13.29</td>
</tr>
</tbody>
</table>

Figure: Data representing real-time amplification data of *L. monocytogenes* with C<sub>t</sub> values (provided in table)

**Sensitivity:** Detectable upto 100-1000 CFU / ml (mg).

**Storage:**
The provided kit has a shelf-life of 6 months when stored at -20°C. Repeated thawing and freezing of PCR reagents should be avoided, as this may reduce the sensitivity. If reagents are to be used multiple times, we recommend storing reagents as aliquots to avoid repeated freeze and thaw. Degradation of sample DNA specimens can also reduce sensitivity of the assay. HiMedia does not recommend using the kit after the expiry date stated on pack.

**Quality Control:**
Each lot of HiMedia’s *Listeria monocytogenes* Detection Kit (Real-time) is assayed for contaminating endonuclease, exonuclease and non-specific DNase activities. Functionally tested in DNA amplification.

**Troubleshooting Guide:**

<table>
<thead>
<tr>
<th>Sr.No.</th>
<th>Problem</th>
<th>Cause</th>
<th>Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>No amplification</td>
<td>Degraded samples</td>
<td>1. Check the integrity of DNA using agarose gel electrophoresis.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>2. Use freshly prepared DNA to ensure the availability of intact template sequence for efficient amplification.</td>
</tr>
<tr>
<td></td>
<td>Error in protocol setup</td>
<td></td>
<td>Verify that the correct reagent volumes, dilutions and storage conditions have been used.</td>
</tr>
</tbody>
</table>

Please refer disclaimer Overleaf.
2. Variability between replicates
   Error in reaction set-up
   Prepare large volume master mix, vortex thoroughly and aliquot into reaction tubes.
   Air bubbles in reaction mix
   Briefly centrifuge reaction samples/plate prior to running on a real-time PCR instrument.
   Pipetting error
   Ct values of replicates can show increased variation due to poor laboratory technique or imprecise pipettes.

3. Amplification in negative control
   Reagents contaminated
   1. Replace all critical solutions
   2. Repeat the analysis of all tests with fresh aliquots of critical reagents.

Safety Information

The Listeria monocytogenes Detection Kit (Real-time) is for laboratory use only, not for drug, household or other uses. Take appropriate laboratory safety measures and wear gloves when handling.

Product Use Limitation & Warranty

HiMedia guarantees the performance of Listeria monocytogenes Detection Kit (Real-time) in the manner described in the product literature. The kit is designed, sold for research and for in vitro purposes only. No claim or representation is intended to provide information for the diagnosis, prevention or treatment of a disease.

All due care and attention should be exercised in the handling of the products. We recommend all users of HiMedia products to adhere to the NIH guidelines that have been developed for recombinant DNA experiments, or to other applicable guidelines.

Technical Assistance

At HiMedia, we pride ourselves on the quality and availability of our technical support. For any kind of technical assistance, mail at mb@himedialabs.com.

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