**Bacillus Cereus Agar Base**

**Intended use**
Recommended as a selective medium for the isolation and enumeration of *Bacillus cereus* from food and clinical samples.

**Composition**

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Gms / Litre</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peptone</td>
<td>1.000</td>
</tr>
<tr>
<td>Mannitol</td>
<td>10.000</td>
</tr>
<tr>
<td>Sodium chloride</td>
<td>2.000</td>
</tr>
<tr>
<td>Magnesium sulphate</td>
<td>0.100</td>
</tr>
<tr>
<td>Disodium hydrogen phosphate</td>
<td>2.500</td>
</tr>
<tr>
<td>Potassium dihydrogen phosphate</td>
<td>0.250</td>
</tr>
<tr>
<td>Sodium pyruvate</td>
<td>10.000</td>
</tr>
<tr>
<td>Bromo thymol blue</td>
<td>0.120</td>
</tr>
<tr>
<td>Agar</td>
<td>15.000</td>
</tr>
<tr>
<td><strong>Final pH (at 25°C)</strong></td>
<td>7.2±0.2</td>
</tr>
</tbody>
</table>

**Directions**
Suspend 20.5 grams in 475 ml purified/distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool to 45-50°C and aseptically add rehydrated contents of 1 vial of Polymyxin B Selective Supplement (FD003) and 25 ml of sterile Egg Yolk Emulsion (FD045). Mix well and pour into sterile Petri plates.

**Principle And Interpretation**

*Bacillus cereus* causes food poisoning due to the consumption of contaminated rice (8,12), eye infections (2) and a wide range of other clinical conditions like abscess formation, meningitis, septicemia and wound infection. *Bacillus cereus* is a known cause of disease mastitis, especially in ewes and heifers among the veterinarians (7). Holbrook and Anderson (4) developed Bacillus Cereus Agar, which is a highly specific and selective medium for the isolation and enumeration of *Bacillus cereus* from foods. It supports the growth of even a small number of *Bacillus cereus* cells and spores in the presence of large number of other food contaminants. The typical colonies of *Bacillus cereus* are crenate, about 5 mm in diameter and have a distinctive turquoise to peacock blue colour surrounded by a good egg yolk precipitate of the same colour. The bacteria do not ferment mannitol and thus there is no change in colour of the indicator dye around the colonies. Addition of polymyxin-B sulphate (3.9) at a final concentration of 100 units per ml of medium is sufficient to make the medium selective for the isolation of *Bacillus cereus*. It suppresses the growth of accompanying bacterial flora. If moulds are suspected in the inoculum, 40 mcg per ml filter-sterilized cycloheximide may be incorporated to suppress the moulds contamination. Some strains of *Bacillus cereus* have very weak egg yolk reaction. Moreover, on this medium *Bacillus cereus* is indistinguishable from *Bacillus thuringiensis*.

Peptone provides and sodium pyruvate improve egg yolk precipitation and enhance sporulation. Bromothymol blue acts as pH indicator to detect mannitol fermentation. For the isolation and enumeration of *Bacillus cereus* in foodstuffs the following method is recommended. Distribute 0.1ml of the homogenized specimen diluted in Peptone Water (M028) onto the surface of the medium. Incubate at 37°C under aerobic conditions for 24-48 hours. Possible growth of contaminants is greatly reduced by incubation for 24 hours. Report the results as the number of *Bacillus cereus* colonies per gram weight of the food sample. Confirmatory tests should be carried out before interpretation.

**Type of specimen**
Food and dairy samples, Clinical sample: eye infections, abscess formation, meningitis, septicemia and wound infection.

**Specimen Collection and Handling**
For food and dairy samples, follow appropriate techniques for sample collection and processing as per guidelines (1,7,11).
For clinical samples, follow appropriate techniques for sample collection and processing as per guidelines (5,6).

After use, contaminated materials must be sterilized by autoclaving before discarding.
**Warning and Precautions**

In Vitro diagnostic use. Read the label before opening the container. Wear protective gloves/protective clothing/eye protection/ face protection. Follow good microbiological lab practices while handling specimens and culture. Standard precautions as per established guidelines should be followed while handling clinical specimens. Safety guidelines may be referred in individual safety data sheets.

**Limitations:**
1. *Bacillus cereus* and *Bacillus thuringiensis* shows identical characteristics and hence difficult to identify.
2. Identification of *Bacillus cereus* is done by colony characteristics and reaction, however further biochemical characteristics should be carried out for confirmation.
3. Some strains of *Bacillus cereus* may show poor growth due to nutritional variations.

**Performance and Evaluation**

Performance of the medium is expected when used as per the direction on the label within the expiry period when stored at recommended temperature.

**Quality Control**

**Appearance**
Cream to greenish yellow homogeneous free flowing powder

**Gelling**
Firm, comparable with 1.5% Agar gel

**Colour and Clarity of prepared medium**
Basal medium : Green coloured clear to slightly opalescent gel. After addition of egg yolk emulsion : Yellowish green coloured opaque gel forms in Petri plates

**Reaction**
Reaction of 4.1% w/v aqueous solution (basal medium) at 25°C. pH : 7.2±0.2

**pH**
7.00-7.40

**Cultural Response**

Cultural characteristics observed with added Polymyxin B Selective Supplement (FD003) and Egg Yolk Emulsion (FD045) after an incubation at 35-37°C for 24-48 hours.

<table>
<thead>
<tr>
<th>Organism</th>
<th>Inoculum (CFU)</th>
<th>Growth</th>
<th>Recovery</th>
<th>Colour of colony</th>
<th>Egg Yolk Reaction</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Bacillus cereus ATCC 10876</em> 50-100</td>
<td></td>
<td>good-luxuriant</td>
<td>&gt;=50%</td>
<td>blue</td>
<td>positive, precipitation</td>
</tr>
<tr>
<td><em>Escherichia coli ATCC 25922 (00013</em>)*</td>
<td>&gt;=10⁴</td>
<td>inhibited</td>
<td>0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Proteus vulgaris ATCC 13315</em> 50-100</td>
<td></td>
<td>good-luxuriant</td>
<td>&gt;=50%</td>
<td>green</td>
<td>negative</td>
</tr>
<tr>
<td><em>Serratia marcescens ATCC 8100</em> 50-100</td>
<td></td>
<td>good-luxuriant</td>
<td>&gt;=50%</td>
<td>yellow-light pink</td>
<td>negative</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>(pigment production is enhanced by incubation at 25-30°C)</td>
<td></td>
</tr>
<tr>
<td><em>Staphylococcus aureus subsp. aureus ATCC 25923 (00034</em>)*</td>
<td>50-100</td>
<td>good-luxuriant</td>
<td>&gt;=50%</td>
<td>yellow</td>
<td>positive, clearing</td>
</tr>
</tbody>
</table>

Key : *Corresponding WDCM numbers.

**Storage and Shelf Life**

Store between 10-30°C in a tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to prevent lump formation due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in dry ventilated area protected from extremes of temperature and sources of ignition Seal the container tightly after use. Use before expiry date on the label. Product performance is best if used within stated expiry period.

**Disposal**

User must ensure safe disposal by autoclaving and/or incineration of used or unusable preparations of this product. Follow established laboratory procedures in disposing of infectious materials and material that comes into contact with clinical
sample must be decontaminated and disposed of in accordance with current laboratory techniques (10,11).

Reference