Corn Meal Peptone Yeast Agar is recommended for the cultivation of fungi.

**Composition**

**Ingredients**

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>Gms / Litre</th>
</tr>
</thead>
<tbody>
<tr>
<td>Corn Meal</td>
<td>20.000</td>
</tr>
<tr>
<td>Dextrose</td>
<td>10.000</td>
</tr>
<tr>
<td>Peptic digest of animal tissue</td>
<td>10.000</td>
</tr>
<tr>
<td>Yeast extract</td>
<td>4.000</td>
</tr>
<tr>
<td>Agar</td>
<td>20.000</td>
</tr>
<tr>
<td>Final pH (at 25°C)</td>
<td>6.5±0.2</td>
</tr>
</tbody>
</table>

**Formula adjusted, standardized to suit performance parameters**

**Directions**

Suspend 64 grams in 1000 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Mix well and pour into sterile Petri plates.

**Principle And Interpretation**

Prospero and Reyes (1) investigated the use of Corn Meal Agar, Soil Extract Agar and Purified Polysaccharide Medium for the morphological identification of *Candida albicans*. Corn Meal Agar is a nutritionally rich medium so it may be also employed for the maintenance of stock cultures of fungi. Corn Meal Peptone Yeast Agar is prepared as per Benjamin (2, 3) for the cultivation of fungi.

The media contain corn meal, which enhances the growth of fungi. Peptic digest of animal tissue and yeast extract provide essential nutrients. Addition of dextrose to the medium supports more luxuriant growth of some fungi as compared to the medium without dextrose, but dextrose supplemented Corn Meal Agar should not be used for chlamydospores production.

**Quality Control**

**Appearance**
Cream to yellow homogeneous coarse powder

**Gelling**
Firm, comparable with 2.0% Agar gel

**Colour and Clarity of prepared medium**
Light amber coloured, opalescent gel forms in Petri plates

**Reaction**
Reaction of 6.4% w/v aqueous solution at 25°C. pH : 6.5±0.2

**pH**
6.30-6.70

**Cultural Response**
M731: Cultural characteristics observed after an incubation at 23 - 27°C for upto 4 days. (For observing Chlamydospore formation:Using a straight wire, make a deep cut in the Corn Meal Agar plate with inoculum. Place a flamed sterile coverslip over the line of inoculum. After incubation, the streaks are examined microscopically, through the coverslip, using low and high power objectives, for chlamydospore formation.)

<table>
<thead>
<tr>
<th>Organism</th>
<th>Inoculum (CFU)</th>
<th>Growth</th>
<th>Chlamydospores Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Aspergillus brasiliensis ATCC 16404</em></td>
<td>50-100</td>
<td>luxuriant</td>
<td>negative</td>
</tr>
<tr>
<td>Candida albicans ATCC 10231</td>
<td>50-100</td>
<td>luxuriant</td>
<td>positive</td>
</tr>
<tr>
<td>Saccharomyces cerevisiae ATCC 9763</td>
<td>50-100</td>
<td>luxuriant</td>
<td>&gt;=70%</td>
</tr>
</tbody>
</table>

Please refer disclaimer Overleaf.
Saccharomyces uvarum 50-100 luxuriant negative >=70%

ATCC 28098

*Key: Formerly known as Aspergillus niger ATCC 16404

Storage and Shelf Life
Store below 30°C in tightly closed container and the prepared medium at 2-8°C. Use before expiry date on the label.

Reference

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